

Implementation of a PET scanner: a guide to reconstruct data from your scanner in STIR

Daniel Deidda, Kris Thielemans

STIR meeting, 3 December 2020

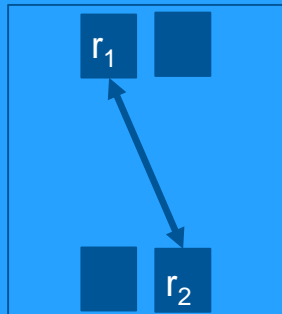
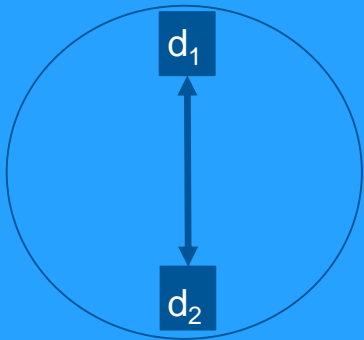
Steps to reconstruct your data

- Read the raw data
- Read data for the Normalisation
- Estimate attenuation
- Align umap and PET image
- Estimate random and scattered events
- Use all the above into the reconstruction

Raw data:

- If interfile, it is easy
- If vendor-specific format:
 - If not public, ask information to the vendor (will need NDA)
 - Collect information to create interfile header with script, implement a scanner object or projData derived class
 - Study the order of detectors and modules

Raw data:



- Binary data is organised as a vector or matrix
- Each element (bin) represents a combination of d_1 , d_2 , r_1 , r_2
- Need to order these bins in a STIR ProjData structure

How?

ProjDataInfoCylindricalNoArcCorr::

get_bin_for_det_pair(Bin& bin, DetectionPositionPair pair)

Raw data: sinogram extraction utility (if vendor raw is a vector/histogram)

```
For (i=0; i<Ntot ; i++) {
```

- Calculate d1, r1, d2, r2 from i
 - proj_info.get_bin_for_det_pos_pair(Bin, pair)
 - Save bin in memory: proj_mem.set_bin_value(bin);
- ```
}
```

```
proj_mem.write_to_file(out_data_filename);
```

## Normalisation: Detector efficiencies

- Similarly to raw data can be saved in vectors (text or binary)
- Need to transform to STIR projData
- `apply_normfactors3D` needs norm terms in a specific order:  $S = t + m * N_t + r * N_t * N_m$
- Important to reorder the values accordingly: create simple utility to manipulate text or binary (ifstream, ofstream)
- Run **`apply_normfactors3D`** with input your STIR-efficiencies file

## Normalisation: Dead-time

- Similarly to efficiencies can be saved in vectors (text or binary)
- Need to transform to STIR ProjData,
- 1 factor for each pair coincidence allowed
- Create look-up table between dead-time index and pair indices
- **ProjDataInfoCylindricalNoArcCorr** Proj\_data\_info
- For(all the sinogram bins){
- Proj\_data\_info.get\_det\_pos\_pair\_for\_bin(pair,bin); (only span 1)
- get pair.pos1 and pair.pos2
- Access table[pos1][pos2]=i and bin.set\_bin\_value(DT\_table[i])
- Proj\_mem.set\_bin\_value(bin)
- }
  
- Proj\_mem.write\_to\_file(out\_data\_filename)

## Attenuation:

Extract the umap from the scanner or use the CT image. If CT image:

- Use **ctac\_to\_mu\_values**, to get a umap ex:  
**ctac\_to\_mu\_values** -o umap.hv -i CT.dcm -j STIR/src/utilities/share/stir/ct\_slopes.json -m mediso -k 511
- Need to subsample ex:  
**zoom\_image** --scaling preserve\_values --template temp.hv zoomed\_umap.hv umap.hv
- Before Calculate\_attenuation\_coefficient we need to register the umap to the PET image
  - Quick recon of PET (no corrections), need to convert to .nii
  - niftyReg: **reg\_aladin** -ref NAC.nii -flo zoomed\_umap.nii -res registration --rigOnly
- Calculate acfsino: **calculate\_attenuation\_coefficients** --PMRT --ACF acfsino zoomed\_umap.hv sino\_template.hs
- **stir\_math** --mult -s normacfsino\_t normsino acfsino
- Also multiply the dead-time similarly to get fullnormsino



## Randoms and scatter:

- Provided with single rates
- Like the efficiencies we need a script to reorder the index (same scripts works)
- Create randoms using:  
**construct\_randoms\_from\_singles** randoms.hs S input\_sino 5
- **Scatter\_and\_recon.sh** script (one for mMR and one for GESigna)
  - All parfiles and scripts are in STIR/example/sample
  - If normsino and/or randoms and/or acfsino is not saved you need to estimate again.
  - Output is total\_additive...hs/s

# Reconstruction: run OSMAPOSL OSMAPOSL.par

$$\lambda_j^{(n+1)} = \frac{\lambda_j^{(n)}}{\sum_i c_{ij}} \sum_i c_{ij} \frac{y_i}{\sum_k c_{ik} \lambda_k^{(n)}} + s_j$$

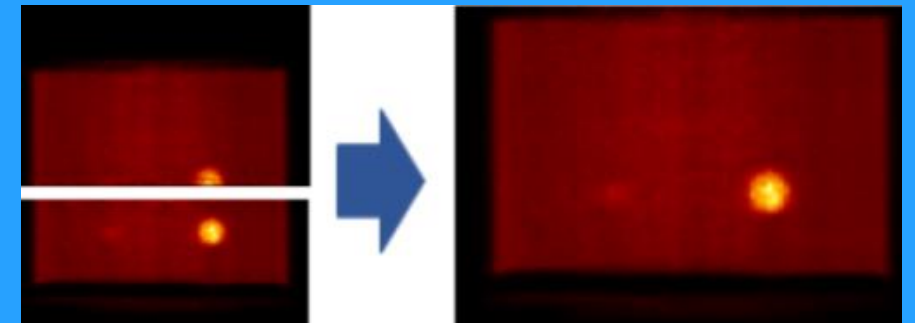
- OSMAPOSLParameters :=
- objective function type:=  
PoissonLogLikelihoodWithLinearModelForMeanAndProjData
- PoissonLogLikelihoodWithLinearModelForMeanAndProjData  
Parameters:=
- input file :=  $\{\text{input\_sino}\}$
- maximum absolute segment number to process := -1
- zero end planes of segment 0 := 1
- projector pair type := Matrix
- Projector Pair Using Matrix Parameters :=
- Matrix type := Ray Tracing
- Ray Tracing Matrix Parameters:=
- number of rays in tangential direction to trace for each bin := 10
- End Ray Tracing Matrix Parameters:=
- End Projector Pair Using Matrix Parameters :=
- recompute sensitivity := 1
- use subset sensitivities := 1
- ; optional. if not set, the subset sensitivities won't be saved
- subset sensitivity filenames:=  $\{\text{recon\_prefix}\}_\text{sens} \%d.hv$
- Bin Normalisation type:=From ProjData
- Bin Normalisation From ProjData :=
- normalisation projdata filename:=  $\{\text{dtnormacfsino}\}$
- End Bin Normalisation From ProjData:=
- additive sinogram :=  $\{\text{additive}\}.hs$
- zoom:= 1
- xy output image size (in pixels) :=  $\{\text{image\_size\_xy}\}$
- z output image size (in pixels) :=  $\{\text{image\_size\_z}\}$
- end  
PoissonLogLikelihoodWithLinearModelForMeanAndProjData  
Parameters:=
- output filename prefix :=  $\{\text{recon\_prefix}\}$
- number of subsets:=  $\{\text{subsets}\}$
- number of subiterations:=  $\{\text{subiter}\}$
- ;start at subiteration number :=1
- Save estimates at subiteration intervals:=  $\{\text{subiter\_interval}\}$
- END :=

**Thank you!**

**Questions?**

## Multiple bed positions:

- Use bed position (BP) info to create a image template for each z offset
- Use `zoom_image` with template to crop umap into multiple BPs
- Created `combine_multiple_bed_position` ex:  
`combine_multiple_bed_positions <out_im_prefix> <num_bed_pos> <iter_num> <bedpost_temp> <image_prefix> <sens_prefix>`
- Iterates over the number of BPs
- Bigger image is created
- Overlapped region is obtained using weighted mean
- `weight=sensitivity`



## Work in progress

- BinNormalisationWithCalibration
- Allows calibration, branching ratio factors
- Radionuclide database
- Decay correction

**Thank you!**